

Original Article

**Effects of pH on Nifekalant-Induced Electrophysiological Change
Assessed in the Langendorff Heart Model of Guinea Pigs**

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Running title: pH and nifekalant

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Abstract

Since information regarding the effects of pH on the extent of nifekalant-induced repolarization delay and torsades de pointes remains limited, we assessed it with a Langendorff heart model of guinea pigs. First, we investigated the effects of pH change from 7.4 to 6.4 on the bipolar electrogram simulating surface lead II ECG, monophasic action potential (MAP), effective refractory period (ERP) and terminal repolarization period (TRP), and found that acidic condition transiently enhanced the ventricular repolarization. Next, we investigated the effects of pH change from 6.4 to 7.4 in the presence of nifekalant (10 μ M) on the ECG, MAP, ERP, TRP and short-term variability (STV) of MAP₉₀, and found that the normalization of pH prolonged the MAP₉₀ and ERP while the TRP remained unchanged, suggesting the increase in electrical vulnerability of the ventricle. Meanwhile, the STV of MAP₉₀ was the largest at pH 6.4 in the presence of nifekalant, indicating the increase in temporal dispersion of repolarization, which gradually decreased with the return of pH to 7.4. Thus, a recovery period from acidosis might be more dangerous than during the acidosis, because electrical vulnerability may significantly increase for this period while temporal dispersion of repolarization remained increased.

Keyword: nifekalant, acidosis, Langendorff, torsades de pointes, monophasic action potential

Introduction

While nifekalant hydrochloride, a class III antiarrhythmic drug, has been clinically used to suppress life-threatening refractory fatal ventricular arrhythmias, the drug was reported to induce excessive QT-interval prolongation, resulting in the onset of torsades de pointes (TdP) ventricular tachyarrhythmia (1-6). These adverse events tended to occur during the recovery period from lethal conditions including severe systemic acidosis, acute renal failure and/or electrolyte disturbance (3-6). Although nifekalant would terminate the ventricular arrhythmia, each organ remained exposed to the low pH blood until the deteriorated systemic circulation was completely recovered (3-7). It is well known that extracellular pH changes can affect the electric activity of cardiomyocytes not only by altering ion flux through pumps and exchangers in the cell membrane (8-9) but also by changing the extent of proton binding to various ion channels (10). Moreover, it has been demonstrated by the patch clamp method in single cells that lowering extracellular pH can weaken the pharmacological effect of some I_{Kr} channel blockers, including dofetilide, azimilide, ibutilide and quinidine (11-12).

Since the information was still lacking regarding the pH effects on the nifekalant-induced electrophysiological change and proarrhythmic risk, we assessed it with the Langendorff heart preparation which allows the control of extracellular conditions (13). We selected guinea-pig hearts for this study, since they have similar ionic channels to those in humans except for I_{to} (14). To precisely analyze the electrophysiological conditions of the ventricle, we measured the monophasic action potential (MAP) and bipolar electrogram simulating surface lead II electrocardiogram (ECG). Furthermore, by adopting an electrical pacing protocol, we measured the MAP duration and effective refractory period (ERP) at a heart rate of 200, 240 and 300 bpm. In this study, we initially analyzed the effects of pH on electrophysiological variables by changing the pH of perfusate from 7.4 to 6.4 and then to 7.4

again; next examined the effects of pH on nifekalant-induced changes in each electrophysiological variable. To precisely estimate the proarrhythmic risk of nifekalant, we examined the temporal dispersion of ventricular repolarization with beat-to-beat analysis method (15).

Materials and Methods

All animal experiments in this study were approved by the Animal Research Committee for Animal Experimentation of Toho University (No.12-51-205) and performed in accordance with the Guidelines for the Care and Use of Laboratory Animals of Toho University and the Japanese Pharmacological Society.

Preparation of Langendorff heart model of guinea pigs and perfusion solution

Male guinea pigs (n=14) weighing 450 to 550 g were anesthetized with sodium pentobarbital (50 mg/kg, i.p.) and heparinized (100 U, i.p.) to protect the heart against microthrombi. The chest was opened at the sternum and the heart was quickly removed. The heart was cannulated through the aorta, and perfused with Krebs-Henseleit solution maintained at $37\pm 0.5^{\circ}\text{C}$ and aerated with a mixture of O_2 (95%) and CO_2 (5%) at a pressure of approximately 60 mmHg and a flow rate of 20 to 30 mL/min by using Langendorff system (Physio-Tech, Tokyo). The Krebs-Henseleit solution contained 118 mM NaCl, 4.7 mM KCl, 11.0 mM glucose, 25 mM NaHCO_3 , 1.2 mM MgSO_4 , 2.5 mM CaCl_2 and 1.2 mM KH_2PO_4 .

A bipolar transcardiac electrogram simulating surface lead II ECG was recorded with two electrodes; one attached onto the apex and the other connected to the base of the heart via a metal cannula. MAP was recorded with a contact electrode placed onto the epicardial surface of the left ventricle. The duration of MAP at a 90% repolarization level (MAP_{90}) was measured. The heart was electrically paced at a cycle length of 300, 250 and 200 ms through a pair of bipolar electrodes contacted onto right ventricle with rectangular pulses of 2-ms duration and approximately twice the threshold voltage to measure the MAP_{90} values; namely, $\text{MAP}_{90(\text{CL}300)}$, $\text{MAP}_{90(\text{CL}250)}$ and $\text{MAP}_{90(\text{CL}200)}$, respectively. ERP of the right ventricle was assessed with programmed electrical stimulation. The pacing protocol consisted of 5 beats of basal stimuli in a cycle length of 300, 250 and 200 ms followed by an

extrastimulus of various coupling intervals to obtain the ERP values; namely, $ERP_{(CL300)}$, $ERP_{(CL250)}$ and $ERP_{(CL200)}$, respectively. The values of MAP_{90} minus ERP at the same pacing cycle length were calculated to estimate the extent of the terminal repolarization period (TRP), which is a reliable marker of electrical vulnerability of the ventricle (16). Starting in the late diastole, the coupling interval was shortened in 5-ms decrements until refractoriness occurred. After the stabilization period, the position and contact pressure of all recording electrodes were not readjusted throughout the experimental period.

Beat-to-beat analysis of MAP_{90} was performed to estimate the extent of temporal dispersion of repolarization (15). Thirty-one consecutive beats of MAP_{90} under sinus rhythm were recorded for each treatment, and Poincaré plots with $MAP_{90(n)}$ versus $MAP_{90(n+1)}$ were prepared. The mean orthogonal distance from the diagonal to the points of the Poincaré plot was determined as short-term variability ($STV = \Sigma |MAP_{90(n+1)} - MAP_{90(n)}| / [30 \times \sqrt{2}]$). The mean distance to the mean of the parameter parallel to the diagonal of the Poincaré plot was determined as long-term variability ($LTV = \Sigma |MAP_{90(n+1)} + MAP_{90(n)} - 2MAP_{90}(\text{mean})| / [30 \times \sqrt{2}]$). The coefficient of variation of MAP_{90} (CV) was calculated with the following equation: standard deviation/mean $\times 100$ (%).

Four types of perfusion solution were prepared according to a previous report (17), namely a normal pH solution (pH 7.4, normal Krebs-Henseleit solution), low pH solution (pH 6.4, modified Krebs-Henseleit solution with $NaHCO_3$ content of 2.5 mM), normal pH solution with drug (10 μ M nifekalant hydrochloride dissolved in normal pH solution), and low pH solution with drug (10 μ M nifekalant hydrochloride dissolved in low pH solution).

Experiment 1: Assessment of baseline electrophysiology under normal and acidic conditions

After equilibration of >30 min of perfusion with the normal pH solution, the preparation was perfused with the low pH solution for 30 min, and then re-perfused with the normal pH solution for 30 min. During each period of perfusion, ECGs under sinus rhythm

and MAP signals at a cycle length of 300, 250 and 200 ms were recorded every 10 min.

ERP of the right ventricle was assessed by programmed electrical stimulation. TRP was also calculated.

Experiment 2: Assessment of electrophysiology under normal and acidic conditions after nifekalant exposure

After equilibration of >30 min of perfusion with the normal pH solution, the preparation was perfused with the normal pH solution for 20 min as the basal control, and then perfused with the low pH solution for 20 min, followed by the low pH solution with 10 μ M of nifekalant for 20 min, and finally perfused with the normal pH solution with 10 μ M of nifekalant for 20 min. At the end of each perfusion, ECG and MAP signals under sinus rhythm, and MAP signals and ERP at a basic cycle length of 300, 250 and 200 ms were recorded. TRP was also calculated.

Drugs and chemicals

The following drugs were purchased: sodium pentobarbital (Kyoritsu Seiyaku Corporation, Tokyo) and sodium heparin (Ajinomoto Pharmaceuticals Co., Ltd. Tokyo). Nifekalant hydrochloride (MW = 441.91) was provided by Nacalai Tesque Inc. (Kyoto). Nifekalant was dissolved in distilled water and small aliquots were added to each perfusion solution to obtain desired final concentration. All other chemicals were commercial products of the available highest quality.

Statistical analysis

Data are presented as mean \pm SE. The differences within a parameter were evaluated by one-way repeated measures analysis of variance (ANOVA). When a p -value was <0.05 by ANOVA, the protocol was judged as having affected the parameter. In this case, the

statistical significance between the treatments was determined by Contrast for mean values comparison, and p -value <0.05 was considered significant.

Results

Experiment 1: Baseline electrophysiology under normal and acidic conditions

The time courses of changes in the MAP_{90} and ERP are summarized in Fig. 1 (n=6) and typical waveforms of the ECG and MAP signals at each treatment are depicted in Fig. 2. Basal control values (C) of the $MAP_{90(CL300)}$, $MAP_{90(CL250)}$ and $MAP_{90(CL200)}$ were 132 ± 8 , 127 ± 7 and 117 ± 6 ms, whereas those of the $ERP_{(CL300)}$, $ERP_{(CL250)}$ and $ERP_{(CL200)}$ were 147 ± 4 , 138 ± 4 and 128 ± 3 ms, respectively. The decrease in pH transiently shortened the $MAP_{90(CL200)}$, and it also shortened the ERP at each basic pacing cycle length. These changes recovered to basal level within 30 min except for the $ERP_{(CL200)}$, which remained shortened during the low pH perfusion as shown in Fig. 1B. These treatments did not affect TRP (data not shown). The morphology of T waves of the ECG under the sinus rhythm significantly changed after pH decreased as depicted in Fig. 2A, indicating a difficulty of precise measurement of QT interval. MAP amplitude remained decreased after lowering pH as depicted in Figs. 2B and 2C. Arrhythmia was not induced by the programmed electrical stimulation during the assessment of ERP at any time point.

Experiment 2: Electrophysiology under normal and acidic conditions after nifekalant exposure

The time courses of changes in the MAP_{90} and ERP are summarized in Fig. 3 (n=8), and those of typical waveforms of the ECG and MAP signals at pH 6.4 and 7.4 are depicted in Fig. 4. Basal control values (C) of the $MAP_{90(CL300)}$, $MAP_{90(CL250)}$ and $MAP_{90(CL200)}$ at pH 7.4 in the absence of nifekalant were 131 ± 7 , 123 ± 6 and 113 ± 5 ms, whereas those of the $ERP_{(CL300)}$, $ERP_{(CL250)}$ and $ERP_{(CL200)}$ were 144 ± 5 , 136 ± 5 and 126 ± 5 ms, respectively. The decrease in pH alone did not significantly affect the MAP_{90} or ERP at 20 min, which was essentially consistent with the results of experiment 1 except for $MAP_{90(CL200)}$ and $ERP_{(CL200)}$.

The difference seems to be explained by the faster adaptation of cellular functions of the preparations in experiment 2 to acidic condition. The addition of nifekalant at pH 6.4 significantly prolonged both the MAP₉₀ and ERP at each pacing cycle length. The increase in pH from 6.4 to 7.4 in the presence of nifekalant further prolonged the MAP₉₀ and ERP at each pacing cycle length except for MAP_{90(CL300)}, which tended to be prolonged but did not achieve statistical significance ($p=0.06$). The increments of MAP₉₀ at a pacing cycle length of 300, 250 and 200 ms were 25 ± 4 , 25 ± 4 , and 20 ± 3 ms, respectively, and there were no significant differences among them. The treatments did not affect TRP (data not shown). The T-wave morphology of ECG under the sinus rhythm significantly changed after pH increased in the presence of nifekalant as depicted in Fig. 4A, also indicating the difficulty of precise measurement of QT interval. MAP amplitude increased with elevating pH in the presence of nifekalant as depicted in Fig. 4B. Arrhythmia was not induced by the programmed electrical stimulation during the assessment of ERP at any time point.

Typical tracings of Poincaré plots are depicted in Fig. 5A and the time courses of the STV, LTV and CV are summarized in Fig. 5B. While lowering pH from 7.4 to 6.4 in the absence of nifekalant did not significantly affect the STV, the administration of nifekalant in low pH solution significantly increased the STV. The normalization of pH in the presence of nifekalant gradually decreased the STV. Although LTV and CV tended to increase at pH6.4 and/or pH7.4 in the presence of nifekalant, these changes did not achieve the statistical significance.

Discussion

This study was designed to assess the effects of pH on the nifekalant-induced repolarization delay and on proarrhythmic risk. The concentration of nifekalant (10 μM) used in this study was determined based on our preliminary experiments in order to better characterize the pharmacological profile of nifekalant, although it is higher than the clinically effective plasma concentration (approximately 0.8 $\mu\text{g/ml}$ [1.8 μM]) in patients with inducible sustained ventricular arrhythmias (18). As depicted in Figs. 2A and 4A, the precise analysis of the effects of each treatment on the repolarization process was difficult in the ECG recordings because of the ambiguity in T-wave terminals, which indirectly supports the utility of currently used MAP recording/pacing technique. Also, the present results indicate that the CV and LTV may be less sensitive in estimating the temporal dispersion of repolarization than the STV as shown in Fig. 5B.

In experiment 1, we analyzed the effects of pH on the electrophysiological variables in the absence of nifekalant. When the preparation was perfused with pH 6.4 solution, phase 2 amplitude of MAP decreased, and MAP duration was transiently shortened as depicted in Fig. 2B. This decrease in phase 2 amplitude of MAP is consistent with previous reports (9,19,20), which has been ascribed to a decrease in plateau-phase Ca^{2+} influx (19,21). Meanwhile, with regard to the shortening of MAP duration, some studies reported that a decrease in extracellular pH shortens the action potential duration (22-25), but the others described opposite results (26,27). It has been reported that under acidic conditions, H^+ directly binds to I_{Kr} and I_{Ca} channels, resulting in their current inhibitions (11,19,20,28). Inhibition of I_{Kr} channels causes the prolongation of MAP duration, whereas inhibition of I_{Ca} channels leads to its shortening. Thus, MAP duration can change in both directions, which depends on the balance in the extent of inhibitions between I_{Kr} and I_{Ca} channels. In addition, acidosis may decrease the ATP production, which opens ATP-dependent K^+ channels, leading

to the shortening of MAP duration (23,29). Therefore, that variability in the response of MAP duration to acidosis among the studies may depend on the differences in methodologies and experimental conditions and/or species differences (29).

In experiment 2, the increase in pH from 6.4 to 7.4 significantly prolonged the MAP₉₀ and ERP in the presence of nifekalant. There have been several clinical case reports describing that nifekalant-induced excessive QT-interval prolongation and TdP during the recovery period from lethal conditions (2-7). In the present experiment with the Langendorff heart preparation, we used pH changes to mimic such pathophysiology in the heart. As summarized in Fig. 3, MAP duration was prolonged less greatly by nifekalant under acidic conditions, which is in accordance with a previous study of guinea-pig ventricular myocytes with the patch clamp method (30), describing that extracellular acidification (pH 6.4) strongly attenuated the effect of nifekalant on the open probability of I_{K1} channel activity when compared with its effect at pH 7.4. As pKa of nifekalant is 7.9, in that paper (30) they speculated that the proportion of ionized to un-ionized form would be greater at pH 6.4 than that at pH 7.4, resulting in a decrease in accessibility to I_{K1} channels via the lipid membrane. Another possible explanation is that an increase in protons may interrupt the binding of nifekalant to I_{Kr} channels (10). Indeed, it is known that there are multiple proton-binding sites in the extracellular domain of the hERG channel, and extracellular protons rapidly and reversely bind to these sites, affecting both hERG activation and deactivation (10). It is possible that the binding of extracellular protons might have attenuated the effects of nifekalant by modulating its binding property. This explanation is indirectly supported by a report that acidic conditions do not affect hERG IC₅₀ of amiodarone, since amiodarone is known to bind to a different site of hERG channels compared with other I_{Kr} blockers whose IC₅₀ was increased under acidic conditions (11). Thus, the attenuation of pharmacological effects of nifekalant under acidic conditions as observed in this study might be due to the decrease in accessibility to I_{K1} channels by increased ionization of nifekalant

and/or the interruption of the binding of nifekalant to I_{Kr} channels by proton binding.

Finally, we investigated the electrical instability of repolarization with the beat-to-beat analysis method, since the Langendorff heart preparation model of guinea pig has been known to be less sensitive in directly detecting the drug-induced TdP (31). The STV of MAP_{90} was the longest at pH 6.4 in the presence of nifekalant. The increase in the STV indicates enhanced “temporal” dispersion and is reported to be closely associated with the onset of early afterdepolarizations and R-on-T-type premature ventricular contractions resulting in the onset of TdP (15). Also, in the present study the MAP_{90} was prolonged, while the TRP remained unchanged by pH normalization at each basic cycle length of 300, 250 and 200 ms in the presence of nifekalant. This observation reflects an increase in electrical vulnerability, in which some cardiomyocytes may have more chance to easily respond to the irregular stimulation from adjacent cells (16). Thus, the most dangerous period for nifekalant-induced TdP would be the recovery period from acidosis, because electrical vulnerability may significantly increase for this one while temporal dispersion of repolarization remained increased.

There are some limitations in this study. In clinical cases with systemic acidosis, multiple pathological factors, including hypoxia, reperfusion injury, superoxide radicals and immune-inflammatory responses, may play important roles in the development of nifekalant-induced QT prolongation and TdP. In order to begin to explore such complex cellular and molecular mechanisms, we assessed the effects of “pH change” on the electrophysiological variables in the hearts in the absence or presence of nifekalant. Based on the current findings, the experiments are now on-going to clarify the effects of the other factors on the nifekalant-induced electrophysiological responses.

In conclusion, the present study with the Langendorff heart model of guinea pigs indicates that MAP recording/pacing technique may be an effective way to better analyze the nifekalant-induced electrophysiological responses in the heart; that pharmacological effects of

nifekalant can be attenuated under acidic conditions; and that nifekalant may induce TdP during the recovery period from systemic acidosis through the increased electrical vulnerability of ventricle and the remaining temporal dispersion of repolarization.

Conflict of interest statement: The authors declare no conflicts of interest.

Acknowledgment: This study was supported in part by Grant-in-aid from the Ministry of Education, Culture, Sports, Science, and Technology of Japan (#25460344, #S1101016), Toho University Joint Research Fund (H23-3, H24-2, H25-3), and the Research Promotion Grant from Toho University Graduate School of Medicine (No.13-01). The authors thank Ms. Misako Nakatani for her technical assistance.

References

- 1 Isomoto S, Konoe A, Cebturion OA, Hayano M, Kaibara M, Hirata T, et al. Electrophysiological effects of MS-551 in humans: a class III antiarrhythmic agent. *Pacing Clin Electrophysiol.* 1995;18:2022-2027.
- 2 Ohashi J, Yasuda S, Miyazaki S, Shimizu W, Morii I, Kurita T, et al. Prevention of life-threatening ventricular tachyarrhythmia by a novel and pure class-III agent, nifekalant hydrochloride. *Journal of Cardiovasc Pharmacol.* 2006;48:274-279.
- 3 Takenaka K, Yasuda S, Miyazaki S, Kurita T, Sutani Y, Morii I, et al. Initial experience with nifekalant hydrochloride (MS-551), a novel class III antiarrhythmic agent, in patients with acute extensive infarction and severe ventricular dysfunction. *Jpn Circ J.* 2001;65:60-62.
- 4 Ando J, Kakishita M, Sakai K, Komura Y, Nishiyama K, Iwabuchi M, et al. Efficacy of nifekalant hydrochloride in the treatment of fatal ventricular arrhythmia in patients with ischemic heart disease. *Int Heart J.* 2005;46:647-656.
- 5 Katoh T, Mitamura H, Matsuda N, Takano T, Ogawa S, Kasanuki H. Emergency treatment with nifekalant, a novel class III anti-arrhythmic agent, for life-threatening refractory ventricular tachyarrhythmias: post-marketing special investigation. *Circ J.* 2005;69:1237-1243.
- 6 Yasuda S, Sawano H, Hazui H, Ukai I, Yokoyama H, Ohashi J, et al. Report from J-PULSE multicenter registry of patients with shock-resistant out-of-hospital cardiac arrest treated with nifekalant hydrochloride. *Circ J.* 2010;74:2308-2313.
- 7 Yoshioka K, Amino M, Morita S, Nakagawa Y, Usui K, Sugimoto A, Matsuzaki A, Deguchi Y, Yamamoto I, Inokuchi S, Ikari Y, Kodama I, Tanabe T. Can nifekalant hydrochloride be used as a first-line drug for cardiopulmonary arrest (CPA)? : Comparative study of out-of-hospital CPA with acidosis and in-hospital CPA without

- acidosis. *Circ J*. 2006;70:21-27.
- 8 Orchard CH, Kentish JC. Effects of changes of pH on the contractile function of cardiac muscle. *Am J Physiol*. 1990;258:C967-981
- 9 Kapur S, Wasserstrom JA, Kelly JE, Kadish AH, Aistrup GL. Acidosis and ischemia increase cellular Ca^{2+} transient alternans and repolarization alternans susceptibility in the intact rat heart. *Am J Physiol Heart Circ Physiol*. 2009;296:H1491-1512.
- 10 Bett GC, Rasmusson RL. Functionally-distinct proton-binding in HERG suggests the presence of two binding sites. *Cell Biochem Biophys*. 2003;39:183-193.
- 11 Lin C, Ke X, Cvetanovic I, Ranade V, Somberg J. The influence of extracellular acidosis on the effect of IKr blockers. *J Cardiovasc Pharmacol Ther*. 2005;10:67-76.
- 12 Lin C, Ke X, Ranade V, Somberg J. Extracellular acidification and hyperkalemia induce changes in HERG inhibition by ibutilide. *Cardiology*. 2008;110:209-216.
- 13 Skrzypiec-Spring M, Grotthus B, Szelag A, Schulz R. Isolated heart perfusion according to Langendorff---still viable in the new millennium. *J Pharmacol Toxicol Methods*. 2007;55:113-126
- 14 Sakaguchi Y, Sugiyama A, Takao S, Akie Y, Takahara A, Hashimoto K. Halothane sensitizes the guinea-pig heart to pharmacological I_{Kr} blockade: comparison with urethane anesthesia. *J Pharmacol Sci*. 2005;99:185-190.
- 15 Thomsen MB, Verduyn SC, Stengl M, Beekman JD, de Pater G, Van Opstal JM, et al. Increased short-term variability of repolarization predicts d-sotalol-induced torsades de pointes in dogs. *Circulation*. 2004;110(16):2453-2459.
- 16 Sugiyama A. Sensitive and reliable proarrhythmia in vivo animal models for predicting drug-induced torsades de pointes in patients with remodelled hearts. *Br J Pharmacol*. 2008;154:1528-1537.
- 17 Garlick PB, Radda GK, Seeley PJ. Studies of acidosis in the ischaemic heart by phosphorus nuclear magnetic resonance. *Biochem J*. 1979;184:547-554.

- 18 Igawa M, Aonuma K, Okamoto Y, Hiroe M, Hiraoka M, Isobe M. Anti-arrhythmic efficacy of nifekalant hydrochloride, a pure class III anti-arrhythmic agent, in patients with healed myocardial infarction and inducible sustained ventricular tachycardia. *J Cardiovasc Pharmacol.* 2002;40:735-742
- 19 Orchard CH, Cingolani HE. Acidosis and arrhythmias in cardiac muscle. *Cardiovasc Res.* 1994;28:1312-1319.
- 20 Du CY, Adeniran I, Cheng H, Zhang YH, Harchi AE, Mcpate MJ, et al. Acidosis impairs the protective role of hERG K(+) channels against premature stimulation. *J Cardiovasc Electrophysiol.* 2010;21:1160-1169.
- 21 Kurachi Y. The effects of intracellular protons on the electrical activity of single ventricular cells. *Pflugers Arch.* 1982;394:264-270.
- 22 Chesnais JM, Coraboeuf E, Sauviat MP, Vassas JM. Sensitivity to H, Li and Mg ions of the slow inward sodium current in frog atrial fibres. *J Mol Cell Cardiol.* 1975;7:627-642.
- 23 Keung EC, Li Q. Lactate activates ATP-sensitive potassium channels in guinea pig ventricular myocytes. *J Clin Invest* 1991;88:1772-1777.
- 24 Sato R, Noma A, Kurachi Y, Irisawa H. Effects of intracellular acidification on membrane currents in ventricular cells of the guinea pig. *Circ Res.* 1985;57:553-561.
- 25 Pacini DJ, Boachie-Ansah G, Kane KA. Modification by hypoxia, hyperkalaemia and acidosis of the cardiac electrophysiological effects of a range of antiarrhythmic drugs. *Br J Pharmacol.* 1992;107:665-670.
- 26 Fry CH, Poole-Wilson PA. Effects of acid-base changes on excitation--contraction coupling in guinea-pig and rabbit cardiac ventricular muscle. *J Physiol.* 1981;313:141-160.
- 27 Kagiya Y, Hill JL, Gettes LS. Interaction of acidosis and increased extracellular potassium on action potential characteristics and conduction in guinea pig ventricular

- muscle. *Circ Res.* 1982;51:614-623.
- 28 Jiang M, Dun W, Tseng GN. Mechanism for the effects of extracellular acidification on HERG-channel function. *Am J Physiol.* 1999;277:H1283-1292.
- 29 Bethell HW, Vandenberg JI, Smith GA, Grace AA. Changes in ventricular repolarization during acidosis and low-flow ischemia. *Am J Physiol.* 1998;275:H551-H561.
- 30 Sato R, Koumi S, Hisatome I, Takai H, Aida Y, Oyaize M, et al. A new class III antiarrhythmic drug, MS-551, blocks the inward rectifier potassium channel in isolated guinea pig ventricular myocytes. *J Pharmacol Exp Ther.* 1995;274:469-474.
- 31 Gerhardy A, Scholtysik G, Schaad A, Haltiner R, Hess T. Generating and influencing Torsades de Pointes--like polymorphic ventricular tachycardia in isolated guinea pig hearts. *Basic Res Cardiol.* 1998;93:285-294.

Figure legends

Fig. 1.

Time courses of the effects of pH change on the electrophysiological variables. (A) Monophasic action potential duration at a 90% repolarization level (MAP₉₀). (B) Effective refractory period (ERP). Extracellular pH was decreased from 7.4 to 6.4, and then increased to 7.4. Data are presented as mean±SE (n=6). **P*< 0.05, ***P*< 0.01 and ****P*< 0.001.

Fig. 2.

Typical tracings of the ECG and MAP, and the summary of MAP amplitude. Decrease in pH to 6.4 from 7.4 followed by an increase to 7.4 changed the morphology of T wave during the sinus rhythm (A), whereas it reduced the amplitude of the MAP during the electrical pacing (B and C). **P*< 0.05 and ***P*< 0.01 compared with pH 7.4 in the absence of nifekalant.

MAP: monophasic action potential.

Fig. 3.

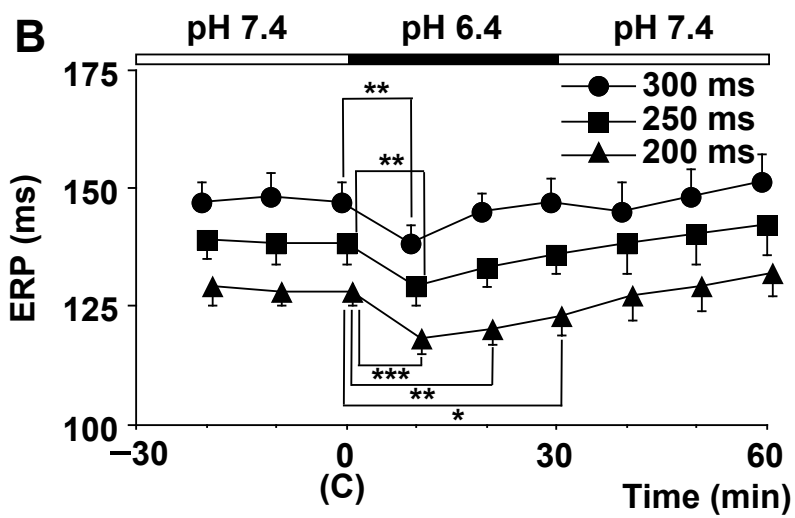
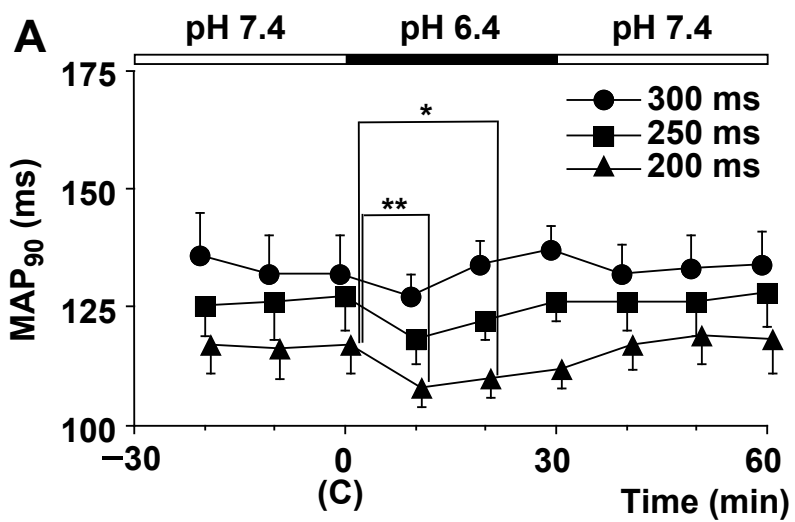
Time courses of the effects of pH change on the electrophysiological variables after nifekalant exposure. (A) Monophasic action potential duration at a 90% repolarization level (MAP₉₀). (B) Effective refractory period (ERP). Extracellular pH was decreased from 7.4 to 6.4, then 10 μM of nifekalant was added, and finally the pH was increased to 7.4 in the presence of nifekalant. Data are presented as mean±SE (n=8). **P*< 0.05, ***P*< 0.01 and ****P*< 0.001.

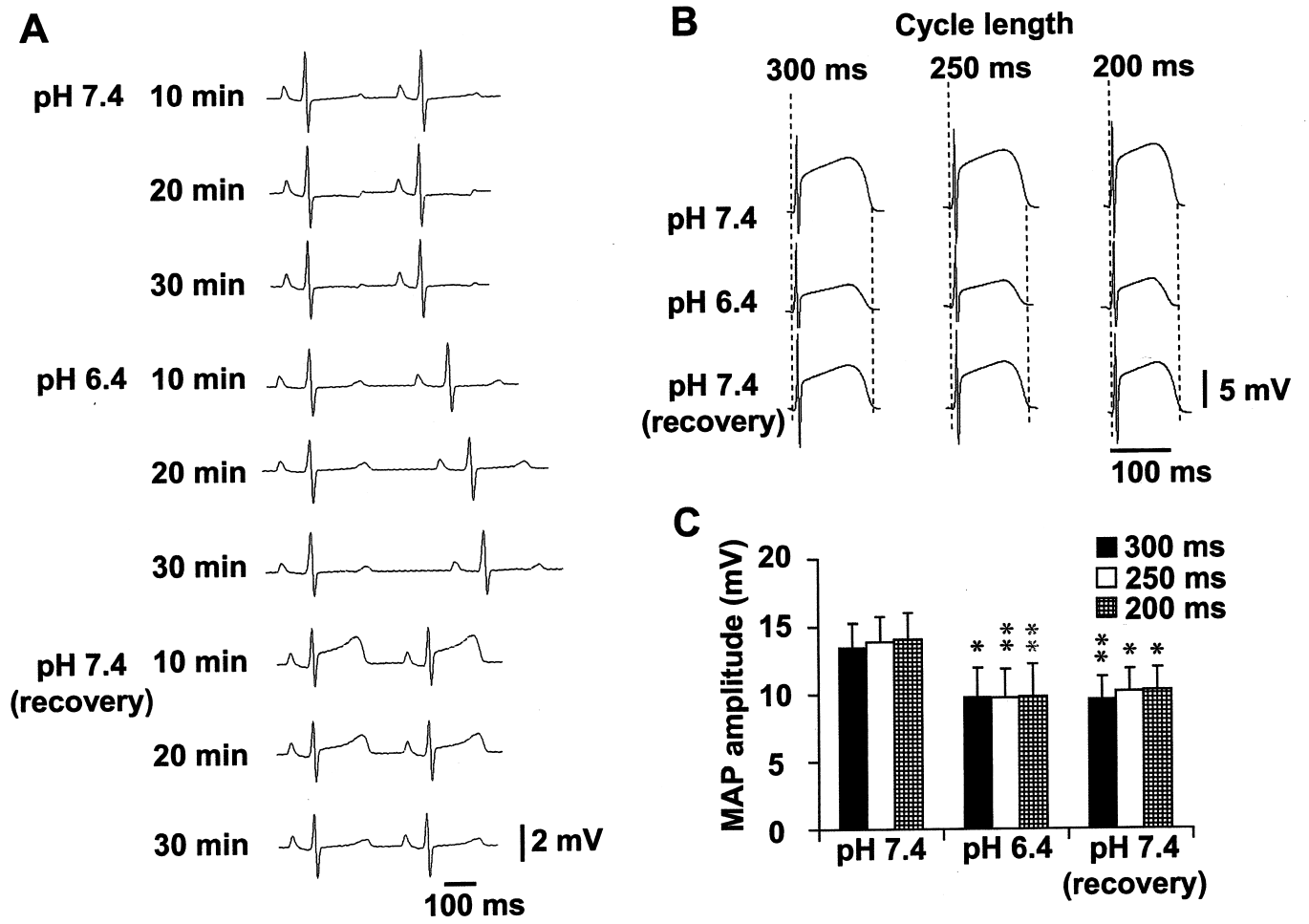
Fig. 4.

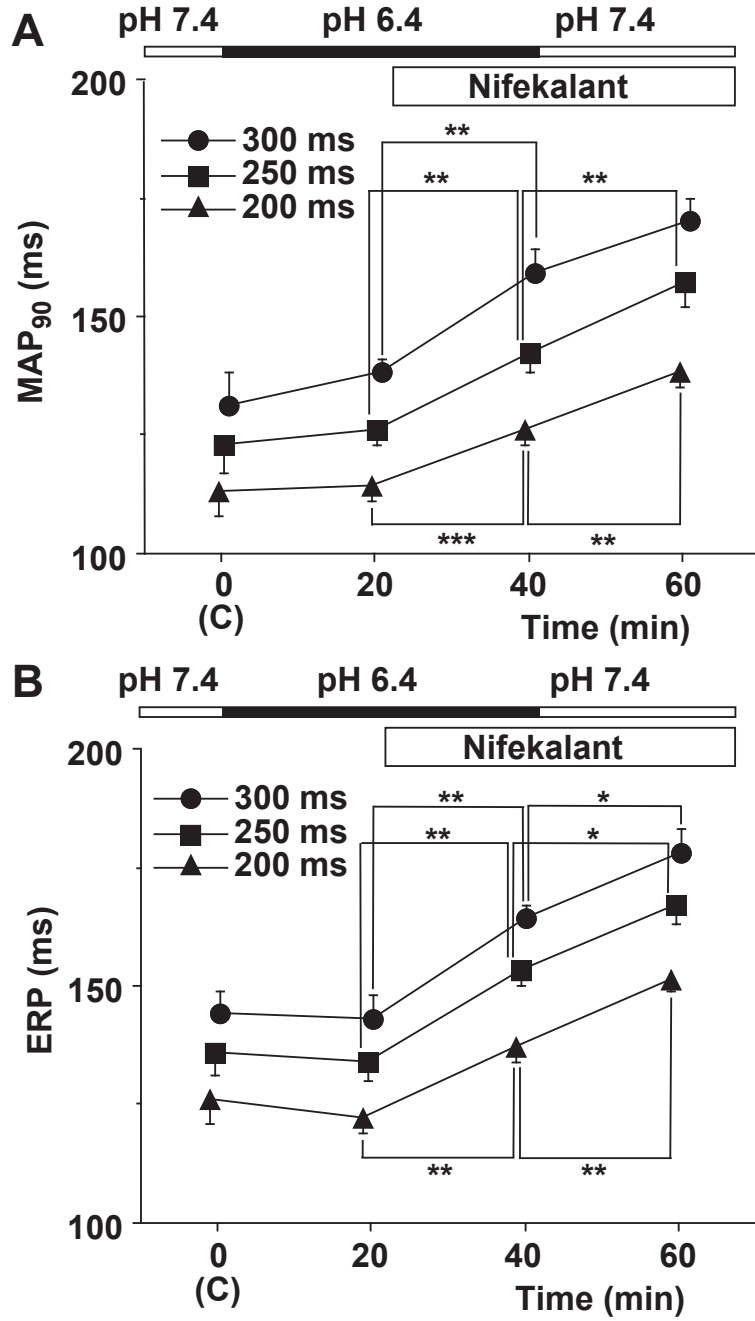
Typical tracings of the ECG (A) and monophasic action potential (B) at pH 6.4 and 7.4. Increase in pH from 6.4 to 7.4 dramatically changed the morphology of T wave during the sinus rhythm (A), whereas it increased the amplitude of the monophasic action potential (B).

Fig. 5

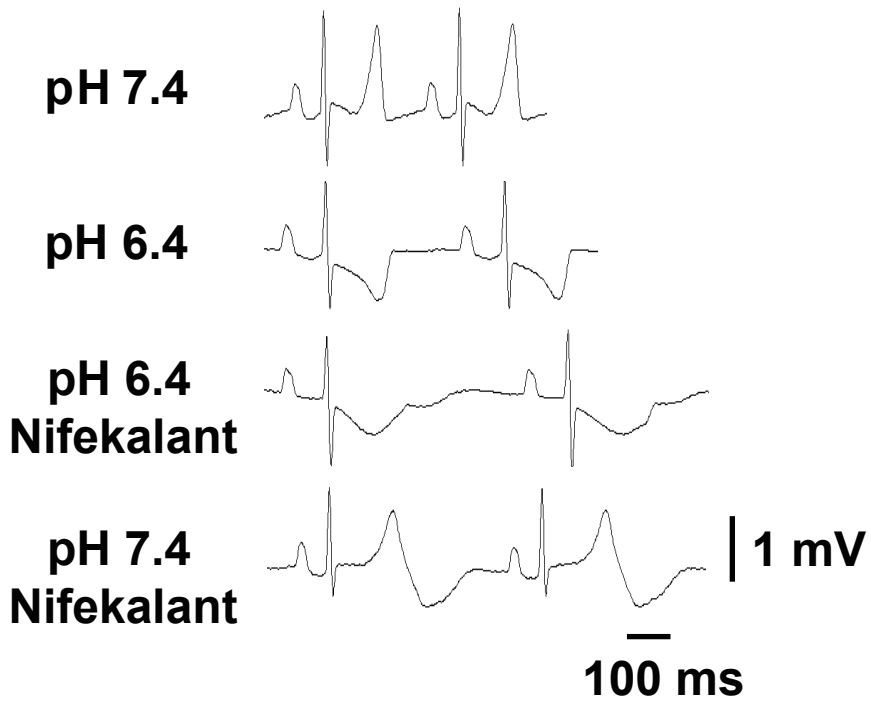
Beat-to-beat analysis of repolarization. (A) Typical tracings of Poincaré plots. (B) Time courses of the STV, LTV and CV. STV: short-term variability; LTV: long-term variability; and CV: coefficient of variation. Data are presented as mean \pm SE (n=8). * $P < 0.05$.







A



B

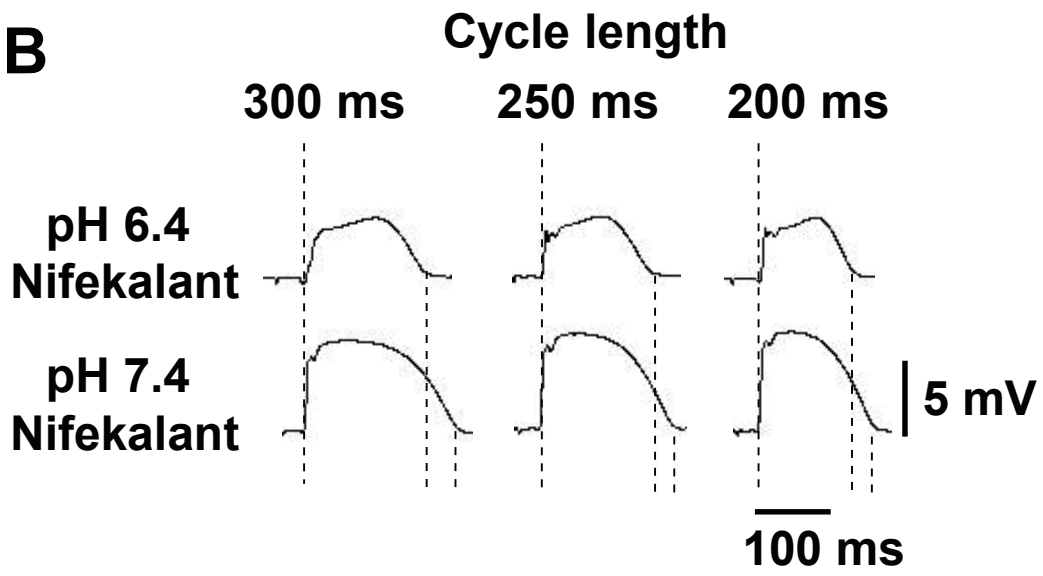


Fig. 5

